



# Revista Mexicana de Oftalmología

[www.elsevier.es/mexoftalmo](http://www.elsevier.es/mexoftalmo)



## ORIGINAL ARTICLE

# Ex vivo human cornea rigidity after UVA/riboflavin induced cross-linking



Alejandro Lichtinger<sup>a,b,\*</sup>, Kátia M. Bottós<sup>a,c</sup>, Jennifer Bernabe-Ko<sup>a</sup>,  
Anne-Catherine Roch-Levecq<sup>d</sup>, Tracy L. Purcell<sup>a</sup>, David J. Schanzlin<sup>a</sup>

<sup>a</sup> Shiley Eye Center, University of California, San Diego, 9415 Campus Point Drive, La Jolla, CA 92093-0946, USA

<sup>b</sup> Instituto de Ciencias Oftalmológicas, Hospital Angeles de las Lomas, Vialidad de la Barranca s/n, Consultorio 735, Huixquilucan, Estado de Mexico CP 52763, Mexico

<sup>c</sup> Department of Ophthalmology, Universidade Federal de São Paulo, Rua Botucatu, 820, São Paulo, SP 04023-062, Brazil

<sup>d</sup> Shiley Eye Center, Department of Biostatistics, University of California, San Diego, 9415 Campus Point Drive, La Jolla, CA 92093-0946, USA

Received 25 October 2012; accepted 21 August 2015

Available online 1 October 2015

## KEYWORDS

Collagen  
cross-linking;  
Riboflavin;  
Ultra-violet light;  
Ex vivo corneal  
measurements;  
Corneal rigidity

## Abstract

**Objective:** The purpose of the current study was to assess changes in human corneal rigidity after treatment with riboflavin and UVA using a new device, the Corneal Displacement Unit (CDU).

**Design:** Experimental study. The CDU was designed to quantitatively measure the effect of cross-linking (CXL) in complete human donor cornea buttons. This apparatus measures the tissue displacement associated to computer-controlled variations in pressure.

**Methods:** Baseline measurements with the CDU were taken from 10 de-epithelialized human corneas. They were exposed to 0.1% riboflavin drops every 2 min for 1 h with exposure to UVA light during the last 30 min after which CDU measurements were taken again. Results were analyzed using a non-parametric test. Significant tests were set at  $P \leq .15$  and non-significant correlation coefficients at  $\rho \leq .15$  to reject abnormal measurements. For each of the 10 corneas, pachymetry values and 1988 CDU measurements were collected before and after treatment.

**Results:** Pressure measurements averaged across the corneas were similar before and after treatment ( $P \geq .99$ ), and were highly correlated (Spearman  $\rho = .88$ ,  $P < .001$ ). Corneal displacement observations after treatment were significantly different from baseline, showing a reduction of .084 microns on average ( $P < .001$ ), and thus demonstrating a reduced displacement in cross-linked corneas.

\* Corresponding author at: Hospital Angeles de las Lomas, Consultorio 735, Vialidad de la Barranca s/n, Col Valle de las Palmas, Huixquilucan, Estado de México CP 52763, Mexico. Tel.: +52 46 9809; fax: +52 46 9808.

E-mail address: [drlichtinger@yahoo.com](mailto:drlichtinger@yahoo.com) (A. Lichtinger).

**PALABRAS CLAVE**

Cross-linking;  
Riboflavina;  
Luz ultravioleta;  
Mediciones en  
corneas ex vivo;  
Rigidez corneal

**Conclusions:** The present study demonstrated a significant increase in the mechanical rigidity of human corneas after CXL. The CDU was able to quantify the mechanical strength of whole corneal buttons ex vivo with minimal damage to the tissue.

© 2015 Sociedad Mexicana de Oftalmología. Published by Masson Doyma México S.A. This is an open access article under the CC BY-NC-ND license (<http://creativecommons.org/licenses/by-nc-nd/4.0/>).

**Cambios inducidos por cross-linking con UVA/riboflavina en la rigidez corneal ex vivo****Resumen**

**Objetivo:** El objetivo de este estudio fue estudiar los cambios en la rigidez corneal tras el tratamiento con Riboflavina y luz ultravioleta (UVA) utilizando un nuevo aparato, la unidad de desplazamiento corneal (CDU).

**Diseño:** Estudio experimental. El CDU fue diseñado para medir cuantitativamente el efecto del cross-linking (CXL) en botones corneales humanos. El instrumento mide el desplazamiento asociado a variaciones en la presión controladas por computadora.

**Métodos:** Tomamos medidas basales con el CDU en 10 corneas tras remover el epitelio. Las corneas fueron expuestas a Riboflavina 0.1%, 1 gota cada 2 minutos durante 1 hora con exposición a UVA durante los últimos 30 minutos, al finalizar se tomaron mediciones con el CDU nuevamente. Los resultados fueron analizados con prueba no paramétrica. La validez se estableció con  $P \leq .15$  y el coeficiente de correlación no significativo en  $\rho \leq .15$  para descartar mediciones anormales. Para cada cornea, valores paquimétricos y 1988 mediciones del CDU fueron tomadas antes y después del CXL.

**Resultados:** Los valores promedio de presión entre las corneas fueron similares antes y después del tratamiento. ( $P \geq 0.99$ ), con una alta congruencia (Spearman  $\rho = 0.88$ ,  $P < 0.001$ ) a diferencia de las mediciones de desplazamiento, donde los valores después de CXL mostraron una reducción promedio de 0.084 micras ( $P < .001$ ); demostrando así un desplazamiento reducido en las corneas tratadas.

**Conclusiones:** Este estudio demuestra un aumento en la rigidez corneal tras tratamiento con CXL. El CDU logro cuantificar el endurecimiento de botones corneales ex vivo con daño mínimo al tejido.

© 2015 Sociedad Mexicana de Oftalmología. Publicado por Masson Doyma México S.A. Este es un artículo Open Access bajo la licencia CC BY-NC-ND (<http://creativecommons.org/licenses/by-nc-nd/4.0/>).

**Introduction**

Corneal cross-linking (CXL) is a novel approach to strengthen the cornea by increasing the mechanical and biochemical stability of stromal tissue. CXL consists in photopolymerization of stromal fibers by the combined actions of a photosensitizing agent (riboflavin) and Ultraviolet-A (UVA), which leads to an increase in the collagen fiber diameter and corneal stiffness.<sup>1,2</sup>

Biomechanical properties of the cornea have been widely studied as a way of addressing corneal changes after CXL, providing crucial information to understand the fundamentals of this treatment. Wollensak and coworkers<sup>3</sup> analyzed corneal rigidity by stress-strain measurements of corneal strips on a microcomputer-controlled biomaterial tester. They found an increase in rigidity, indicated by rise in stress, of more than 300% in treated human corneas. Later, Kohlhaas et al.<sup>4</sup> used corneal biomechanical properties to describe the localized effect of CXL in the anterior portion of strips of human and porcine corneas.

Until recently, the study of tissue mechanical properties has relied on methods such as tensile tests, which require the samples to be cut. To better ascertain the effects of the CXL process, it is necessary to utilize techniques that are capable of evaluating biomechanical properties within complete corneas. The aim of the present study was to assess changes in human corneal rigidity after treatment with riboflavin and UVA irradiation. For this purpose we designed a new device, the Corneal Displacement Unit (CDU), to quantitatively measure the effect of CXL in complete human donor cornea buttons. This apparatus measures the tissue displacement associated to computer-controlled variations in pressure. Results obtained from this study may further expand our understanding of CXL process and its potential role in our therapeutic armamentarium.

**Materials and methods**

The CDU was designed with the help of the bioengineering department at our university; once the final model was built,

several test runs with human corneal buttons demonstrated stability of the measurements; corneas without exposure to riboflavin or UVA light were maintained in the artificial chamber and measurements repeated twice during the next hour, these preliminary data taught us that as long as we kept a stable pachymetry throughout the experiment the displacement remained steady. Taking this into consideration we designed the present study to assess the effect of CXL in corneal displacement with the CDU.

Ten research grade corneas not suitable for corneal transplantation were obtained from the San Diego Eye Bank. The post-mortem time of the human eyes was less than 48 h. Corneas that had prior surgery, scarring or pathology that could affect corneal clarity or hydration were not accepted for the study.

We removed the corneal epithelium from 10 corneas, took baseline measurements with the CDU, and then exposed the corneas to 0.1% isotonic riboflavin drops (10 mg riboflavin-5-phosphate in 10 ml dextran T-500 20%) every 2 min for 1 h with exposure to UVA light during the last 30 min after which CDU measurements were taken again.

Pachymetry measurements were performed in all corneas immediately after removal of the epithelium, before UVA exposure and before the second CDU measurement to ensure a stable pachymetry between readings ( $\pm 40 \mu\text{m}$ ).

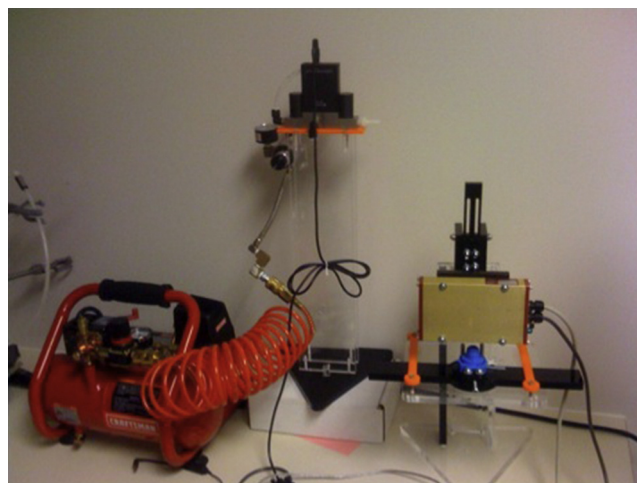
### UVA irradiation

UVA exposure was performed with a Vega LED-based UV emitter (CSO Srl. Firenze, Italy) assembled and tested according to the manufacturers instructions. The Vega LED is preset to emit radiation of 370 nm wavelength at an intensity of  $3.0 \text{ mW/cm}^2$ ; a UV meter was used to check the UV emission power at the treatment plane before exposure. Exposure was carried out for 30 min with a 10 mm treatment diameter as per the manufacturer's protocol.

### Corneal Displacement Unit

The CDU consists of a computer regulated piezo-electric pressure transducer that is connected to an artificial anterior chamber via semi-rigid surgical tubing filled with balanced salt solution (Fig. 1). Ex vivo human corneal tissue was then mounted onto the artificial anterior chamber and a watertight seal was obtained. This system allowed for precise computer control of the internal pressure within the artificial anterior chamber using LabView (National Instruments Corporation, Austin, TX).

A HeNe laser emitter and solid-state detector unit with fixed placement were both mounted above the apex of the corneal tissue. After internal calibration of the laser, the tissue was placed at an appropriate distance from the laser such that the reflected beam was within the detectors range to allow for measurements of the physical displacement of the corneal apex. These measurements were obtained using LabView and were correlated with the corresponding pressure applied by the pressure transducer within the artificial anterior chamber. We placed the corneas in pressure cycles that range from 0.1 to 1.0 PSI, which corresponds to an intraocular pressure of 5.6 mmHg to 56 mmHg, with a frequency of 6 cycles per minute for 10 min.



**Figure 1** Corneal Displacement Unit: a computer regulated piezo-electric pressure transducer is connected to an artificial anterior chamber via semi-rigid surgical tubing filled with balanced salt solution. This system allowed for precise computer control of the internal pressure within the artificial anterior chamber using LabView (National Instruments Corporation, Austin, TX).

We performed preliminary testing in 5 corneas which were placed in the CDU for a full hour without the CXL procedure; these corneas showed pressure and corneal displacement measurements that were stable throughout the timeline validating the accuracy and baseline measurements for our study. In this study we compared pretreatment and post-CXL CDU measurements in each cornea tested.

### Statistical analysis

For each of the 10 corneas, pachymetry values and 1988 CDU measurements were collected before and after treatment. Stability of measurements before and after treatment was examined for each cornea using non-parametric tests, i.e., Wilcoxon matched pairs test, and Spearman correlation. Significant tests were set at  $P \leq .15$  and non-significant correlation coefficients at  $\rho \leq .15$  to reject abnormal measurements. All measurements were then averaged across all corneas retained for analysis and analyzed using non-parametric tests to assess any change in displacement post-CXL treatment from preoperative CDU measurements.

### Results

We were able to perform all the measurements in 9 out of 10 corneas; results from corneal button # 1 were discarded due to abnormal values caused by a leak from the artificial anterior chamber. This leak was likely caused by a small diameter scleral ring. Another 2 corneal button measurements were discarded after statistical analysis due to skewed observations between the data collected before and after CXL. The computer captured too many data points at the highest pressure values on the untreated corneal buttons and too many points at the lowest pressure values once the corneal buttons were CXL; these data had a correlation near to 0 and therefore could not be utilized in our study.

**Table 1** Mean (SD), median (range) measurements of IOP in PSI for 10 human corneas before and after CXL treatment.

	Cornea	Before CXL	After CXL	P values*
1.	Mean (SD)	.625 (.270)	.335 (.270)	<.001
	Median (range)	.660 (.09–1.0)	.199 (1.0–1.0)	
	Skewness	–0.330	1.169	
	Kurtosis	–1.135	–0.053	
2.	Mean (SD)	.622 (.268)	.623 (.270)	0.867
	Median (range)	.660 (.06–1.0)	.660 (.02–1.0)	
	Skewness	–0.310	–.336	
	Kurtosis	–1.144	–1.134	
3.	Mean (SD)	.623 (.269)	.625 (.269)	0.599
	Median (range)	.660 (.06–1.0)	.660 (.08–1.0)	
	Skewness	–0.321	–.331	
	Kurtosis	–1.137	–1.133	
4.	Mean (SD)	.624 (.270)	.625 (.273)	0.859
	Median (range)	.660 (.06–1.0)	.670 (.06–1.0)	
	Skewness	–0.340	–.344	
	Kurtosis	–1.130	–1.149	
5.	Mean (SD)	.624 (.271)	.624 (.273)	0.396
	Median (range)	.670 (.10–1.0)	.680 (.09–1.0)	
	Skewness	–0.341	–.349	
	Kurtosis	–1.144	–1.169	
6.	Mean (SD)	.624 (.270)	.721 (.296)	<.001
	Median (range)	.660 (.08–1.0)	.790 (.07–1.1)	
	Skewness	–0.326	–.547	
	Kurtosis	–1.141	–0.987	
7.	Mean (SD)	.625 (.269)	.623 (.269)	0.947
	Median (range)	.660 (.02–1.0)	.660 (.05–1.1)	
	Skewness	–0.328	–.336	
	Kurtosis	–1.136	–1.128	
8.	Mean (SD)	.622 (.267)	.621 (.270)	0.872
	Median (range)	.660 (.05–1.0)	.660 (.10–1.0)	
	Skewness	–0.318	–.318	
	Kurtosis	–1.138	–1.157	
9.	Mean (SD)	.609 (.303)	.552 (.290)	<.001
	Median (range)	.650 (.08–1.0)	.550 (.10–1.0)	
	Skewness	–0.213	–.023	
	Kurtosis	–1.403	–1.408	
10.	10 Mean (SD)	.619 (.268)	.625 (.267)	0.456
	Median (range)	.650 (.06–1.0)	.660 (.10–1.0)	
	Skewness	–0.285	–.330	
	Kurtosis	–1.161	–1.145	

\* Wilcoxon matched pairs test.

Tables 1 and 2, respectively, show IOP and corneal displacement measurements for 10 human corneas before and after CXL treatment. Pressure measurements averaged across the 7 remaining corneas were similar before and after treatment ( $P \geq .99$ ), and were highly correlated (Spearman  $\rho = .88$ ,  $P < .001$ ). Corneal displacement observations after treatment were significantly different from baseline, showing a reduction of  $.084 \mu\text{m}$  on average ( $P < .001$ ), and thus demonstrating a reduced displacement in cross-linked corneas (Table 3, Fig. 2).

## Discussion

In the current study, we demonstrated an increase in mechanical stability by the reduction on corneal displacement of *ex vivo* human corneas after treatment with riboflavin and UVA. The CDU provides reproducible displacement values of the cornea with minimal damage to the tissue. This new technique may potentially validate and compare new treatment strategies by evaluating changes in corneal stiffness caused by CXL.

**Table 2** Mean (SD), median (range) for corneal displacement measurements in microns for 10 human corneas before and after CXL treatment.

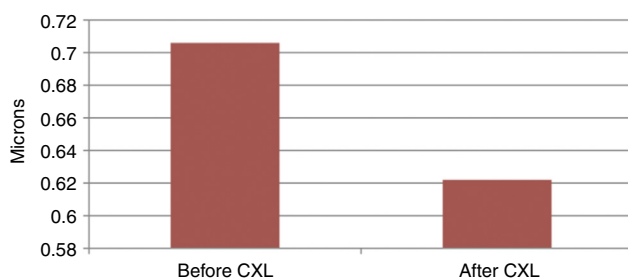
	Cornea	Before CXL	After CXL	Mean difference	P values*
1. <sup>E</sup>	Mean (SD)	.111 (.003)	.093 (.002)	-.018 (.003)	<.001
	Median (range)	.112 (.082-.114)	.199 (.076-.095)		
2.	Mean (SD)	.099 (.002)	.107 (.003)	.008 (.003)	<.001
	Median (range)	.098 (.098-.110)	.106 (.105-.118)		
3.	Mean (SD)	.078 (.003)	.073 (.001)	-.005 (.003)	<.001
	Median (range)	.079 (.068-.082)	.073 (.064-.075)		
4.	Mean (SD)	.117 (.001)	.092 (.012)	-.025 (.012)	<.001
	Median (range)	.118 (.115-.123)	.073 (.071-.125)		
5.	Mean (SD)	.116 (.002)	.092 (.001)	-.026 (.002)	<.001
	Median (range)	.116 (.102-.119)	.091 (.088-.095)		
6. <sup>E</sup>	Mean (SD)	.107 (.002)	.104 (.001)	-.003 (.002)	<.001
	Median (range)	.107 (.095-.120)	.103 (.101-.106)		
7.	Mean (SD)	.108 (.003)	.053 (.001)	-.055 (.003)	<.001
	Median (range)	.107 (.105-.123)	.053 (.051-.055)		
8.	Mean (SD)	.067 (.002)	.082 (.001)	.015 (.001)	<.001
	Median (range)	.067 (.061-.071)	.082 (.082-.086)		
9. <sup>E</sup>	Mean (SD)	.109 (.002)	.109 (.001)	-.001 (.002)	.250
	Median (range)	.109 (.107-.122)	.109 (.108-.110)		
10.	Mean (SD)	.121 (.001)	.122 (.001)	.001 (.001)	<.001
	Median (range)	.121 (.117-.124)	.122 (.120-.125)		

\* Wilcoxon matched pairs test.

<sup>E</sup> Excluded from final analysis due to unstable IOP measurements.**Table 3** IOP and corneal displacement measurements averaged across the 7 remaining corneas before and after CXL treatment.

Cornea	Before CXL	After CXL	Mean difference	<i>P</i> values*
<i>IOP</i>				
Mean (SD)	4.36 (1.60)	4.37 (1.32)	.01 (.80)	.99
Median (range)	4.54 (.59–6.72)	4.49 (.68–4.92)	.05 (.09–1.80)	
<i>Displacement</i>				
Mean (SD)	.706 (.007)	.622 (.014)	–.084 (.014)	<.001
Median (range)	.704 (.684–.732)	.615 (.597–.664)	–.09 (.87–.68)	

\* Wilcoxon matched pairs test.

**Figure 2** Corneal Displacement before and after CXL.

CXL has emerged as a promising alternative to intra-stromal implants, penetrating or lamellar keratoplasty in the treatment of keratoconus and post-refractive ectasia.<sup>5-7</sup> Therefore, new technologies to assess the effects of CXL are needed. The majority of experimental studies<sup>3,4,8</sup> used tensile measurements in order to assess stress-strain properties after CXL. Although these techniques provide important

biomechanical data, they require the cornea to be sectioned, which alters the properties of the corneal button functioning as an entire unit. Some of these experimental problems include the following: the collagen fibers at the edge of these sliced samples are fragmented, thus reducing the predictability of the measurements; the segmentation of the corneal buttons leaves them susceptible to changes in hydration that can further modify the biomechanical properties of the tissue. Furthermore, because these samples must be clamped in the jaws of the device, the edges are submitted to compressive tension that may alter the final results. In contrast, the CDU, used in the present study, allows us to analyze the response of the whole cornea as a unit. There is no need for segmenting the tissue since the whole button is mounted on the artificial chamber where it stays in place during the whole experiment. This also allows us to maintain constant pressure behind the cornea during the time between measurements. Characteristics of this CDU may optimize biomechanical measurements and serve as a model for further applications.



Corneal biomechanical analysis has enhanced our understanding of the CXL process. In 2003, Wollensak<sup>3</sup> first described the results of mechanical measurements of human corneal strips after CXL. An increase in rigidity of 328.9% was observed after treatment, with a corresponding increase of 4.5 in Young's modulus. The same biomaterial measuring device was used by Kohlhaas et al.<sup>4</sup> to evaluate stress strain characteristics of the anterior and posterior portions of the cornea. Their results provided further evidence that the effect of CXL is greater in the anterior stroma.<sup>9–13</sup> Recently, this type of measurements was also used to evaluate the CXL effect after modifications of the standard CXL technique. Rigidity analysis was used to compare CXL effects on corneas with and without de-epithelialization<sup>14</sup> and it was demonstrated that corneas with intact epithelium had a decreased stiffening effect. Similar studies were performed by McCall et al.,<sup>8</sup> but in all of these studies, and in contradistinction with the CDU described in the present manuscript, cutting the corneas into segments was necessary to allow for the measurement of biomechanical data.

The need for a non-destructive technique to assess corneal stiffness and biomechanical properties is soaring; therefore other investigators, including Duppset et al.<sup>15</sup> and Mattson et al.<sup>16</sup> have explored other potential techniques that may allow the analysis of corneal rigidity in whole corneal buttons. A technique named Globe Expansion Method (GEM)<sup>16</sup> was tested in rabbit eyes with the purpose of measuring the behavior of cross-linked tissue to elevated intra-ocular pressure; in this study, corneas submitted to riboflavin and UVA had a reduced expansion compared with control corneas. Another non-invasive technique named Surface Wave Elastometry<sup>15,17</sup> attempts to assess biomechanical properties by measuring the velocity of the wave propagation along the long axes of the collagen fibrils. The authors tested, in vitro, the changes in corneal elasticity after CXL and found that surface wave velocity increased after treatment, thus assuming that the cornea was strengthened by increased stiffness. Although using different techniques from the one described in our paper, the results obtained by these non-destructive techniques seem to concur with our findings.

In conclusion, we demonstrated a significant increase in the mechanical rigidity of human corneas after CXL, providing further evidence of the effect of riboflavin/UVA on corneal tissue. We have developed novel equipment, which we named the Corneal Displacement Unit, to quantify the mechanical strength of whole corneal buttons *ex vivo* with minimal damage to the tissue. This new device could be helpful in comparing different CXL protocols, new riboflavin formulations and delivery methods of cross-linking components. We believe this and other related devices could further expand our knowledge within this developing field of cross-linking.

## Ethical responsibilities

**Protection of people and animals.** The authors declare that the procedures followed were in accordance with the regulations of the responsible Clinical Research Ethics Committee and in accordance with those of the World Medical Association and the Helsinki Declaration.

**Confidentiality of data.** The authors declare that no patient data appears in this article.

**Right to privacy and informed consent.** The authors must have obtained the informed consent of the patients and/or subjects mentioned in the article. The author for correspondence must be in possession of this document.

## Funding

No endorsement of any kind received to conduct this study/article.

## Conflict of interest

The authors declare no conflict of interest.

## References

1. Wollensak G, Spoerl E, Seiler T. Riboflavin/ultraviolet-A-induced collagen crosslinking for the treatment of keratoconus. *Am J Ophthalmol.* 2003;135:620–7.
2. Wollensak G, Wilsch M, Spoerl E, et al. Collagen fiber diameter in the rabbit cornea after collagen crosslinking by riboflavin/UVA. *Cornea.* 2004;23:503–7.
3. Wollensak G, Spoerl E, Seiler T. Stress-strain measurements of human and porcine corneas after riboflavin/UVA induced cross-linking. *J Cataract Refract Surg.* 2003;29:1780–5.
4. Kohlhaas M, Spoerl E, Schilde T, et al. Biomechanical evidence of the distribution of cross-links in corneas treated with riboflavin and ultraviolet A light. *J Cataract Refract Surg.* 2006;32:279–83.
5. Vinciguerra P, Albè E, Trazza S, et al. Refractive, topographic, tomographic, and aberrometric analysis of keratoconic eyes undergoing corneal cross-linking. *Ophthalmology.* 2009;116:369–78.
6. Raiskup-Wolf F, Hoyer A, Spoerl E, et al. Collagen crosslinking with riboflavin and ultraviolet-A light in keratoconus: long-term results. *J Cataract Refract Surg.* 2008;34:796–801.
7. Wollensak G. Crosslinking treatment of progressive keratoconus: new hope. *Curr Opin Ophthalmol.* 2006;17:356–60.
8. McCall AS, Kraft S, Edelhauser HF, et al. Mechanisms of corneal tissue cross-linking in response to treatment with topical riboflavin and long-wavelength ultraviolet radiation (UVA). *Invest Ophthalmol Vis Sci.* 2010;51:129–38.
9. Mazzotta C, Balestrazzi A, Traversi C, et al. Treatment of progressive keratoconus by riboflavin-UVA-induced cross-linking of corneal collagen: ultrastructural analysis by Heidelberg Retinal Tomograph II in vivo confocal microscopy in humans. *Cornea.* 2007;26(4):390–7.
10. Bottós KM, Dreyfuss JL, Regatieri CV, et al. Immunofluorescence confocal microscopy of porcine corneas following collagen cross-linking treatment with riboflavin and ultraviolet A. *J Refract Surg.* 2008;24:S715–9.
11. Spoerl E, Mrochen M, Sliney D, et al. Safety of UVA-riboflavin cross-linking of the cornea. *Cornea.* 2007;26:385–9.
12. Esquenazi S, He J, Li N, et al. Immunofluorescence of rabbit corneas after collagen cross-linking treatment with riboflavin and ultraviolet A. *Cornea.* 2010;29:412–7.
13. Wollensak G, Lomdina E, Dittert DD, et al. Wound healing in the rabbit cornea after corneal collagen cross-linking with riboflavin and UVA. *Cornea.* 2007;26(5):600–5.

14. Wollensak G, Iomdina E. Biomechanical and histological changes after corneal crosslinking with and without epithelial debridement. *J Cataract Refract Surg.* 2009;35:540–6.
15. Dupps WJ Jr, Netto MV, Herekar S, et al. Surface wave elastometry of the cornea in porcine and human donor eyes. *J Refract Surg.* 2007;23(1):66–75.
16. Mattson MS, Huynh J, Wiseman M, et al. An in vitro intact globe expansion method for evaluation of cross-linking treatments. *Invest Ophthalmol Vis Sci.* 2010;51:3120–8.
17. Rocha KM, Ramos-Esteban JC, Qian Y, et al. Comparative study of riboflavin-UVA cross-linking and “flash-linking” using surface wave elastometry. *J Refract Surg.* 2008;24(7):S748–51.